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<p>(21) International Application Number: PCT/US89/02619</p> <p>(22) International Filing Date: 15 June 1989 (15.06.89)</p> <p>(30) Priority data: 207,298 15 June 1988 (15.06.88) US</p> <p>(71) Applicants: WHITEHEAD INSTITUTE FOR BIOMEDICAL RESEARCH [US/US]; Nine Cambridge Center, Cambridge, MA 02142 (US). MEDICAL RESEARCH COUNCIL [GB/GB]; 20 Mount Pleasant, London WIN 4AL (GB).</p> <p>(72) Inventors: YOUNG, Richard, A. ; 5 Sawmill Brook Road, Winchester, MA 01890 (US). YOUNG, Douglas ; 44 Lawnclose Ruislip, Middlesex HA4 6ED (GB).</p> <p>(74) Agents: GRANAHAN, Patricia et al.; Hamilton, Brook, Smith & Reynolds, Two Militia Drive, Lexington, MA 02173 (US).</p>		<p>(81) Designated States: AT (European patent), BE (European patent), CH (European patent), DE (European patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent).</p> <p>Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>
<p>(54) Title: STRESS PROTEINS AND USES THEREFOR</p> <p>(57) Abstract</p> <p>Stress proteins and their use to immunize an individual against a nonviral infection or to induce immune tolerance in an individual, as well as a method of immunizing an individual by administering a selected stress protein and a method of inducing immune tolerance in an individual by administering a selected stress protein.</p>		

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STRESS PROTEINS AND USES THEREFORDescriptionBackground of the Invention

05 Although the function of stress proteins is not
entirely clear, it appears that some participate in
assembly and structural stabilization of certain cellular
and viral proteins, and their presence at high
concentrations may have an additional stabilizing effect
during exposure to adverse conditions. Neidhardt, F.C.
10 and R.A. VanBogelen, In: Escherichia coli and Salmonella
typhimurium, Cellular and Molecular Biology, (eds.
Neidhardt, F.C., Ingraham, J.L., Low, K.B., Magasanik, B.
Schaechter, M. and Umbarger, H.E. (Am. Soc. Microbiol.,
Washington, D.C.), pp. 1334-1345 (1987); Pelham, H.R.B.
15 Cell, 46:959-961 (1986); Takano, T. and T. Kakefuda,
Nature, 239:34-37 (1972); Georgopoulos, C. et al., New
Biology, 239:38-41 (1972). Phagocytic host cells produce
a hostile environment for foreign organisms, and the
ability to produce stress proteins has been implicated in
20 the survival of bacterial pathogens within macrophages
Christman, M.F. et al., Cell, 41:753-762 (1985).

Mycobacterium (M.) tuberculosis and Mycobacterium
(M.) leprae are the etiologic agents of tuberculosis and
leprosy, respectively. These diseases afflict 20-30
25 million people and continue to present a significant
global health problem. Joint International Union Against
Tuberculosis and World Health Organization Study Group,
Tubercle, 63:157-169 (1982); Bloom, B. and T. Godal, Rev.
Infect Dis. 5:765-780 (1983). To develop more effective

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that most of the antigens identified with monoclonal antibodies are involved in the T-cell response to mycobacterial infection or vaccination in mice and in humans. Limiting dilution analysis indicates that 20% of the mycobacterial-reactive CD4⁺ T lymphocytes in mice immunized with M. tuberculosis recognize a single protein, the 65-kDa antigen. Kaufman, S.H.E. et al., Eur J. Immunol., 17:351-357 (1987).

Summary of the Invention

10 The present invention relates to stress proteins and methods of modulating an individual's immune response, either to a pathogen or to his or her own cells, such as occurs in autoimmune diseases. In particular, it relates to the use of such stress proteins as a "vaccine" in
15 immune prophylaxis therapy, which results in an induction or enhancement of immune response to a selected pathogen and as an immunotherapeutic agent in treatment of autoimmune diseases, which results in a decrease of an individual's response to his or her own cells. In immune
20 prophylaxis, stress proteins are administered to prevent or reduce the effects in an individual of a pathogen, which can be any virus, microorganism or other organism or substance (e.g., a toxin or toxoid) which causes disease. In preventing or reducing adverse effects of
25 nonviral pathogens (e.g., bacteria, mycobacterial) according to the method of the present invention, an individual's immune response to the nonviral pathogen's stress protein(s) is induced or enhanced through the administration of a vaccine which includes the pathogen's
30 stress protein(s) and, generally, an adjuvant.

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protein can be administered in multiple doses over time in order to induce immune tolerance against an autoimmune disease such as rheumatoid arthritis.

Brief Description of the Drawings

05 Figure 1 is a graphic representation of the homologies between mycobacterial antigens and known stress proteins. Figure 1A is a representation of sequence similarity between portions of the M. tuberculosis 71-kDa antigen (residues 1-204; TB 71kDa) and the E. coli DnaK protein (residues 430-469). Figure 1B is a
10 representation of sequence similarity between portions of the M. tuberculosis 65-kDa antigen (residues 1-540; TB 65kDa) and the E. coli GroEL protein (residues 1-547).

15 Figure 2 is a comparison of the amino acid sequence of the human P1 protein (573 residues) and the amino acid sequence of the groEL protein (547 residues).

20 Figure 3 is a comparison of the amino acid sequence of the human P1 protein (573 residues), which is a homolog of groEL protein, and the amino acid sequence of the 65kDa M. leprae protein (540 residues).

 Figure 4 is a comparison of the amino acid sequence of the human P1 protein (573 residues), which is a homolog of the groEL protein, and the amino acid sequence of the 65kDa M. tuberculosis protein (540 residues).

25 Detailed Description of the Invention

 The present invention is based on the observation that stress proteins are among the major antigens available for presentation to T lymphocytes and may be common immune targets in a broad spectrum of infectious

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Exp. Med., 165:1430-1435 (1987)) and the malarial parasite Brugia malayi (Selkirk, M.E. et al., J. Cell Biochem., 12D:290 (1988)). Similarly, homologues of GroEL have been found among antigens involved in the

05 immune response to Salmonella typhimurium and Coxiella. Vodkin, M.H. and J.C. Williams, J. Bacteriol., 170:1227 (1988). The presence of stress proteins among major

10 immune targets in a variety of human pathogens is support for the idea that the stress response may be a general component of infection and that stress proteins should be considered among candidates for subunit vaccines. All

15 organisms respond to heat by inducing synthesis of heat shock proteins (hsps), which are a group of proteins. This response is the most highly conserved genetic system known and has been shown to occur in every organism,

20 including microorganisms, plants and animals, investigated to date. Many of the characteristics of the response are common to all organisms and the hsps are among the most highly conserved proteins known. For

25 example, hsp90 family and hsp70 family proteins are present in widely diverse organisms. The proteins in each family--even in such diverse organisms--show approximately 50% identity at the amino acid level and at the nonidentical residues, exhibit many similarities.

30 Several of the proteins induced by heat are also induced by a variety of other stresses. The hsps or a closely related/similar protein are present in all organisms at normal temperatures and have been shown to have key functions in normal cell metabolism. Lindquist, S. and E.A. Craig, Ann. Rev. Genet., 22:631-677 (1988). Because the stress response is common to prokaryotes and

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In view of the involvement of proteins of M. tuberculosis and M. leprae in humoral and cell-mediated immune responses and to establish the functions of these proteins in the mycobacterial cell, the DNA
05 encoding several of the M. tuberculosis and M. leprae antigens have been sequenced. It has been demonstrated, as a result, that many of these mycobacterial protein antigens exhibit striking sequence similarity to known stress-induced proteins. Three of the M. leprae and
10 two of the M. tuberculosis protein antigens studied have been shown to exhibit striking sequence similarity to known stress proteins. For reasons discussed in the Exemplification, it is concluded that two of the M. leprae and two of the M. tuberculosis antigens are
15 homologues of the E. coli DnaK and GroEL proteins.

In experimental mice, immunization with mycobacterial lysates elicits antibody responses to at least six M. tuberculosis protein antigens and a similar number of M. leprae protein antigens. Monoclonal anti-
20 bodies specific for these proteins have been used to isolate clones from λ gt11 DNA expression libraries of M. tuberculosis and M. leprae. The sequence of the DNA clones revealed that mycobacterial hsp70 (alias 70 kDa antigen) and hsp60 (alias 65 kDa antigen, groEL) were the
25 major targets of the murine antibody response to both M. tuberculosis and M. leprae. Two additional hsp's, an 18 kDa member of the small hsp family and a 12 kDa homologue of groES, were found among the M. leprae and M. tuberculosis antigens. Young, D.B., et al., Proc. Natl. Acad. Sci. USA, 85:4267-4270 (1988); Shinnick, T.M., et al., Nuc. Acids Res., 17:1254 (1989).

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some fraction of the human T cell clones were shown to recognize one or more of the mycobacterial stress proteins.

05 Stress Proteins are Immune Targets in Non-viral Infections

The observation that stress proteins are important targets of the immune response to mycobacterial infection and the knowledge that the major stress proteins are conserved and abundant in other organisms suggested that stress proteins are likely to be immune targets in many non-viral infections. Indeed, that is now clearly the case. Antigens from a wide variety of infectious agents have been identified as members of stress protein families. The major stress protein antigen recognized by antibodies in bacterial infections is hsp60. "Common antigen", an immunodominant protein antigen long known to be shared by most bacterial species, turns out to be hsp60. Shinnick, T.M., et al., Infect. Immun., 56:446 (1988); Thole, J.E.R., et al., Microbial Pathogenesis, 4:71-83 (1988). Stress proteins have also been identified as immune targets in most major human parasite infections. Bianco, A.E., et al., Proc. Natl. Acad. Sci. USA, 83:8713 (1986); Nene, V., et al., Mol. Biochem. Parasitol., 21:179 (1986); Ardesir, F., et al., EMBO J., 6:493 (1987); Hedstrom, R., et al., J. Exp. Med., 165:1430 (1987); Selkirk, M.E., et al., J. Cell Biochem., 12D:290 (1988); Engman, D.M., et al., J. Cell Biochem., 12D: Supplement, 290 (1988); Smith, D.F., et al., J. Cell Biochem., 12D:296 (1988). Antibodies to hsp70 have been identified in the sera of patients

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of years. A number of different proteins present in synovial membranes have been proposed to be the cross-reactive rat antigen, but were later discounted as procedures for the purification of these proteins improved. van Eden, W., et al., Proc. Natl. Acad. Sci. USA, 82:5117-5120 (1985); Holoshitz, J., et al., Science, 219:56-58 (1983). The M. tuberculosis antigen recognized by the arthritogenic T cells was recently shown to be a 65 kDa protein (van Eden, W., et al., Nature, 331:171 (1988), which has now been shown to be hsp60 (see the Exemplification). Using a combination of truncated recombinant 65 kDa proteins and peptides, a nine amino acid epitope of hsp60 has been identified as the minimum stimulatory sequence for arthritogenic T cell clones in proliferation assays. Now that it is clear that some arthritogenic T cells recognize the mycobacterial hsp60, it is quite possible that the rat autoantigen is also hsp60.

The results obtained in the adjuvant arthritis model led investigators to determine whether T lymphocytes from human rheumatoid arthritis patients also recognize mycobacterial antigens. These investigators have found not only that patients with rheumatoid arthritis have T cells that recognize M. tuberculosis antigens, but that these T cells have diverse phenotypes. Substantial proliferative responses to mycobacterial extracts are observed with uncloned T cells (predominantly CD4⁺) from both synovial infiltrates and peripheral blood, although responses are generally greater in synovial infiltrates. Abrahamson, T.G., et al., Scand. J. Immunol., 7:81-90 (1978); Holoshitz, J., et al., Lancet ii, 305-306 (1986).

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self stress protein determinants; and observations that stress responses are induced by viral infection and by cell transformation, all suggest a model of immune surveillance in which self-reactive T cells provide a first line of defense against infection and transformation by recognizing and helping to eliminate stressed autologous cells, as well as cells infected with intracellular bacteria. The pool of lymphocytes that recognize conserved stress protein determinants might be induced during establishment of natural microbial flora on the skin and in the gut, and maintained by frequent stimulation by bacteria and viruses as well as other stressful stimuli encountered during a normal lifetime. This model is attractive because it provides a way in which the immune system could exploit the existence of conserved epitopes in stress proteins to respond immediately to antigenically diverse pathogens and cellular changes, producing an initial defense that need not await the development of immunity to novel antigens.

Stress protein induction occurs in eukaryotic cells following infection by diverse viruses in vitro. Collins, P.L., and Hightower, L.E., J. Virol., 44:703-707 (1982); Nevins, J.R., Cell, 29:913-939 (1982); Garry, R.F., et al., Virology, 129:391-332 (1988); Khandjian, E.W. and Turler, H., Mol. Cell Biol., 3:1-8 (1983); LaThangue, N.B., et al., EMBO J., 3:267-277 (1984). CTL that recognize these neo-antigens could limit the spread of virus by killing infected cells, possibly before substantial amounts of mature virus are assembled, and by secreting the lymphokine γ -interferon. Pestka, S., in: Methods Enzymol., Interferons, Part A., Vol. 79, Academic

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proteins are constitutively expressed in normal cells, although at lower levels than in stressed cells, the potential for autoreactivity is ever-present. Normal cells may escape destruction by expressing only
05 substimulatory levels of stress protein determinants on their surfaces. In addition, stress proteins may only be processed and presented during stress, and it may be relevant that many stress proteins have altered
intracellular locations during stress. Finally, immune
10 regulatory networks may prevent activation of autoreactive T cells under normal conditions. The regulatory constraints required by this system might occasionally break down, perhaps during stress caused by bacterial or viral infections, leading to autoimmune
15 disease. Rheumatoid arthritis may be such a disease.

Modulation of Immune Response

The precise relationship between stress proteins and the host immune response to infection is as yet undefined. When cells are subjected to a variety of
20 stresses, they respond by selectively increasing the synthesis of a limited set of stress proteins. Some stress proteins, including the products of dnaK and groEL, are major constituents of the cell under normal growth conditions and are induced to even higher levels
25 during stress. Lindquist, S., Annu. Rev. Biochem., 55: 1151-1191 (1986); Neidhardt, F.C. and R.A. VanBogelen, In Escherichia coli and Salmonella Typhimurium, Cellular and Molecular Biology, (eds. Neidhardt, F.C., Ingraham, J.L. Low, K.B. Magasanik, B. Schaechter, M. and Umberger,
30 H.E.) Am. Soc. Microbiol., Washington, D.C., pp. 1134-1345 (1987). It has now been demonstrated that stress-

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response (and, thus, reducing the pathogen's effects) can be used.

First, because the nonviral pathogen's stress proteins are distinguishable from those of the host, it is possible to induce an immunoprophylactic response specific to the pathogen's stress proteins. This can be carried out by administering a vaccine which includes all or a portion (e.g., sufficient sequence to have the desired stimulatory effect on immune response) of the pathogen's stress protein or of another protein having an amino acid sequence sufficiently similar to that of the stress protein sequence to stimulate the immune response to the pathogen. Alternatively, highly conserved stress protein determinants, such as those shown to be recognized by a variety of T cells, can be administered as a type of "general" vaccine. In either case, the immune response to the stress protein sequence will be increased and effects of the nonviral pathogen will be reduced (decreased, prevented or eliminated).

Second, it is also possible to induce or enhance the immune surveillance system or response which is directed to stressed host cells. This is described further in the context of enhancing immune response in those instances in which the pathogen (e.g., a virus, transforming agent) does not have (express) its own stress proteins (i.e., stress proteins distinguishable from host stress proteins).

The vaccine administered to induce or enhance immune response to nonviral pathogens includes a stress protein of the pathogen against which an immune response is desired, a portion of that protein of sufficient size to

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or any other substances or changes in condition which induce the stress response in the individual being treated. (This can also be employed in conjunction with the vaccine, described previously, administered to
05 enhance immune response to a stress protein-producing pathogen.) It is known that increased levels of stress proteins are produced in many types of cancer cells. Enhancement of the immune surveillance system, as described, can be used to facilitate destruction and/or
10 to prevent progression or establishment of cancer cells.

The method of the present invention can also be used to modify or modulate an individual's response to his or her own cells (e.g., as in autoimmune diseases). There are at least two ways in which the present invention can
15 be used immunotherapeutically. First, stress proteins, such as heat shock protein (hsp) 70 and hsp60, are known to be involved in autoimmune disease. It is, thus, possible to turn down an individual's response to "self" by administering the appropriate stress protein(s) in
20 such a manner that the individual becomes more tolerant of the protein. Second, because it is known that the immune response in autoimmune diseases is to stress proteins, it is possible to selectively inhibit or interfere with the ability of immune cells which normally
25 interact with such proteins to do so. This can be done, for example, by administering monoclonal antibodies that bind to specific T cell receptors and delete or disable such cells. Alternatively, rather than knocking out immune cells, the stress response in all cells can be
30 turned down by administering a drug capable of reducing a cell's ability to undergo the stress response. For

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(1980). DNA sequences were determined for both strands of DNA. Computer analysis of sequences with UWGCC programs was as described by Devereux, J., et al. Nucleic Acids Res., 12:387-395 (1984).

- 05 Immunoblot Analysis. Escherichia coli strain TGI was transformed with the following plasmids by standard procedures (Maniatis, T., et al., Molecular Cloning, A Laboratory Manual (Cold Spring Harbor Lab., Cold Spring Harbor, NY) (1982), with selection for ampicillin resistance: pND5, a derivative of pBR325 containing the E. coli groE genes (Jenkins, A.J., et al., Mol. Gen. Genet. 202:446-454 (1986); pUC8 (Vic?, J., Gene, 19:259-268 (1982); pUC8 with insert DNA for λ gt11 clone Y3178 (M. leprae 65-kDa antigen, Young, R.A., et al., Nature, (London) 316:450-452 (1985)) ligated in the EcoRI site.

- Overnight cultures of E. coli strains in Luria-Bertani (LB) medium were centrifuged and resuspended in isotonic phosphate-buffered saline at a cell density corresponding to an absorbance of 20 at 60 nm. An equal volume of sample buffer containing 2% (wt/vol) polycrylamide gels in the presence of NaDodSO_4 was added, and, after heating on a boiling water bath for 2 min, 5-ml samples were electrophoresed on 12% (wt/vol) polycrylamide gels in the presence of NaDodSO_4 . Blots were prepared by electrophoretic transfer of the proteins to a nitrocellulose membrane, and binding of monoclonal antibodies was assayed with a peroxidase-conjugated secondary antibody as described. Young, D.B., et al., Infect. Immun., 55:1421-1425 (1987).

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TABLE Mycobacterial protein antigens

		Recognized by human T cells	Subjected to sequence analysis	Homology with known proteins
<u>Protein, kDA</u>		<u>-----</u>		
05	M. tuberculosis			
	71	+	+	DnaK
	65*	+	+	GroEL
	38	+	-	-
	19	+	+	None
10	14	+	-	-
	12	ND	-	-
	M. leprae			
	70	ND	-	DnaK
	65	+	+	GroEL
15	36	+	-	-
	28	+	-	-
	18	+	+	Plant Hsp
	12	ND	-	-

20 Mycobacterial protein antigens, their recognition by human T cells, and homology of the deduced mycobacterial protein sequences to known proteins are summarized.

ND, not determined; +, yes; -, no

* Includes data derived from study of the 65-kDA antigens of M. bovis BCG (bacillus Calmette-Guerin), which is identical to the M. tuberculosis 65-kDA antigen.

+ A. S. Mustafa, J. R. Lamb, D. Young and R. A. Young, unpublished data.

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The Mycobacterial 65-kDa antigen. The 65-kDa antigens of M. tuberculosis and M. leprae are involved in the human T-cell response to mycobacterial infection (Table). Genes encoding these proteins have been

05 isolated (Husson, R.N. and Young, R.A., Proc. Natl. Acad. Sci., USA, 84:1679-1683 (1987); Young, R.A., et al., Nature, (London) 316:450-452 (1985)) and sequenced (Shinnick, T.M., J. Bacteriol., 169:1080-1088 (1987);

10 Mehram, V., et al., Proc. Natl. Acad. Sci., USA, 83:7013-7017 (1986)), revealing that the amino acid sequences of the 65-kDa antigens of M. tuberculosis and M. leprae are 95% identical. These protein sequences exhibit no significant sequence similarity to proteins in the GenBank database.

15 Identification of these proteins was based on the observation that some monoclonal antibodies directed against the mycobacterial 65-kDa antigens cross-react with an E. coli protein of 60kDa. E. coli cells transformed with the plasmid pND5 (Sanger, F., et al., Proc. Natl. Acad. Sci., USA, 74:5463-5467 (1977), which

20 contains the E. coli groE genes, had been shown to accumulate large amounts of the 60-kDa protein. A comparison of the mycobacterial 65-kDa protein sequences with those determined for E. coli groEL (C. Woolford, K. Tilly, C. Georgopoulos, and R.H., unpublished data)

25 revealed the extent of the sequence similarity as shown in Fig. 1B.

The 60-kDa Gro EL protein is a major stress protein in E. coli. Lindquist, S., Annual Rev. Biochem., 55:1151-1191 (1986); Nature, 333:330-334 (1988). There

30 is some evidence that the mycobacterial 65-kDa proteins

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CLAIMS

1. A vaccine comprising all or a portion of a selected stress protein or all or a portion of a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of the stress protein.
05
2. A vaccine of Claim 1 in which the stress protein is a mycobacterial stress protein or a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of the mycobacterial stress protein.
10
3. A composition for use as an agent to induce immune tolerance, comprising a selected stress protein.
4. A composition for use in treating an autoimmune disease, comprising all or a portion of a selected stress protein or all or a portion of a protein having an amino acid sequence sufficiently homologous to the amino acid sequence of the stress protein.
15
5. A composition of Claim 4 for treating rheumatoid arthritis.
20
6. A vaccine for use in enhancing in an individual the immune response to a nonviral pathogen, comprising all or a portion of a stress protein of the nonviral pathogen against which the enhanced response is desired.
25

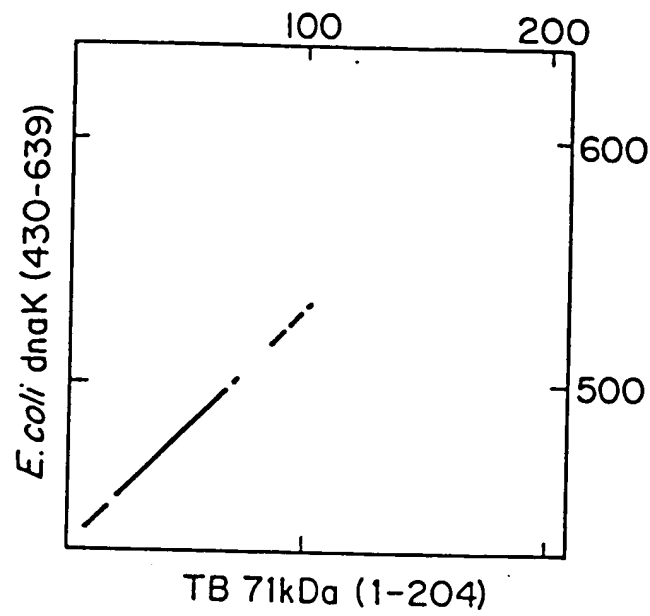
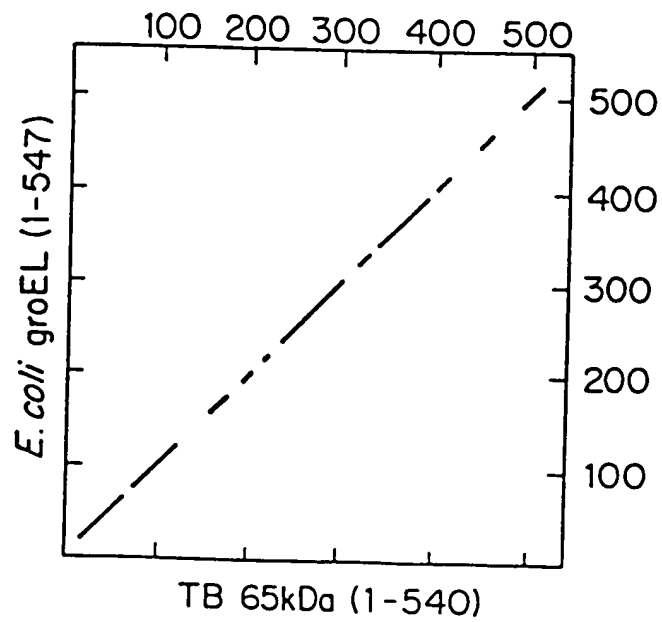
*Fig. 1A**Fig. 1B*

FIGURE 2

1	10	20	30	40	50	60	70
HUMPI	MLRLPTVFRQMRPVSRLAPHLTRAYAKDVKFGADARALMLQGVLLADAVAVTMGPKGRTVIEEQSWGS						
GROEL	-----MA-----AKDVKFGNDARVKMLRGVNVLDADVKTGLGPKGRNVVLDKSFGE						
71	80	90	100	110	120	130	140
HUMPI	PKVTKDGVTVAKSIDLKDKNIGAKLVQDVANNTNEEAGDGTATVLAARSIAKEGFEKISKGANPVEI						
GROEL	PTITKDGVSVAIEPEPKFENMGAMQVKEVASKANDAAGDGTATVLAQAIIITEGLKAVAAAGMNPMDL						
141	150	160	170	180	190	200	210
HUMPI	RRGVMLAVDAVIAELKKQSKPVTTPEEIAQVATISANGDKEIGNI						
GROEL	RGIDKAVTAAVEELKALSVPCSDSKAIAQVGTISANSDETVGKLEAMDKVKGEGVITVEDGTGLQDE						
211	220	230	240	250	260	270	280
HUMPI	LEIIEGMMKFDGRGYISPYFINTSKGQKCEQDAYVLLSEKKISSIQSIVPALEIANAHKPLVIAEDVDG						
GROEL	LDVVEGMQFDRGYLSPYFINKPETGAVELESPPFILLADKKISNIREMLPVLAVAKAGKPLLI AEDVEG						
281	290	300	310	320	330	340	350
HUMPI	EALSTLVNRLKVLQVAVAKAPGFGDNRKNQLKDMAIATGGAVFGEGLTNLEDVQPHDLGKVGVEVIV						
GROEL	EALATAVVNTIRGIVKVAAVKAPGFGDRRKAMLQDIATLTGTVISEE-IGMELEKATLEDLGQAKRVVI						
351	360	370	380	390	400	410	420
HUMPI	TKDDAMLLKGKDKAQIEKRIQEIIEQLDVTTSEYEKEKLNRLAKLSDGVAVLVKGGTSDVEVNEKKDR						
GROEL	NKDTTTIIDGVGEEAAIQGRVAQIRQIQIEEATSDYDREKLQERVAKLGGVAVIKVGAATEVEMKEKKAR						

FIGURE 2 CONT'D

421	430	440	450	460	470	480	490
HUMPI	VTDALNATRAAVEEGIVLGGGCALLRQ	IPALDLSLTPANEDQKIGIEIIKRTLKIPAMTI	AKNAGVEGSLI				
GROEL	VEDALHATRAAVEEGVAGGGVALIRV	ASKLADLRQ	NEDQNVSSSL-RAMEAPLRQIVLNCGEPSVV				
491	500	510	520	530	540	550	560
HUMPI	VEKIMQSSSEVGYDAMAGDFVNMVEKGIIDPTKVV	RTALLDAAGVASLLTTAEVVVVTEIPKEKDPGMGA					
GROEL	ANTVKGGDGNYGYNAAATEEYGNMIDMGILDPTK	VT	RSALQYAA	SVAGL	MITTECMVTDLPKND-AADLGA		
561	570						
HUMPI	MGGMGG--GMGGGMF						
GROEL	AGGMGGMGGMGM-						

Total score = 4667, 5 breaks
 276 identities out of 545 possible matches between residues

25 random runs
 Alignment score = 65.34 SD Standard deviation = 18.94 Mean = 3429.48

FIGURE 3

1	10	20	30	40	50	60	70
HUMP1	MLRLPTVFRQMRPVSRLAPHLTRAYAKDVKFGADARALMLQGVDLLADAVATMGPKGRTVIEQSWGS						
ML65K	M-----AKTIAYDEEARRGLEGLNSLADAVKVTGLGPKGRNVVLEKKWGA						
71	80	90	100	110	120	130	140
HUMP1	PKVTKDGVTVAKSIDLKDKYKNIGAKLVQDVANNTNEEAGDGTATTATVLARSIAKEGFEKISKGANPVEI						
ML65K	PTITNDGVSIAKEIELEDPEYKIGAEELVKEVAKKTDVAGDGTATTATVLAQALVKEGLRNVAAGANPLGL						
141	150	160	170	180	190	200	210
HUMP1	RRGVMLAVDAVIAELKKQSKPVTTPEEIAQVATISANGDKKEIGNIISDAMKKVGRKGVITVKDGTKLTNDE						
ML65K	KRGIEKAVDKVTETLLKDAKEVETKEQIAATAAISA-GDQSIGDLIAEAMDKVGNEGVIIVEESNTFGLQ						
211	220	230	240	250	260	270	280
HUMP1	LEIIEGMMKFDRGYISPYFINTSKGQKCEFQDAYVLLSEKKISSIQSIVPALEIANAHKPLVIAEDVDG						
ML65K	LELTEGMRFDKGYISGYFVTDARERQEAVLEEPYILLVSSKVSTVKDLLPLEKVIQAGKSLLIIAEDVEG						
281	290	300	310	320	330	340	350
HUMP1	EALSTLVNRLKVGLQVVAVKAPFGDNRKNQLKDMAIATGGAVFGEGLTLNLEDVQPHDLGKVGEVIV						
ML65K	EALSTLVNKKIRGTFKSVAVKAPFGDNRKAMLQDMAILTGAQVISEE-VGLTLENTDLSLLGKARKVVM						
351	360	370	380	390	400	410	420
HUMP1	TKDDAMLLKKGDKDAQIEKRIQEIIEQLDVTTSEYEKEKELNERLAKLSDGVAVLKVGGTSDVEVNEKKDR						
ML65K	TKDETTIVEGAGDTDAIAGRVAQIRTEIENSDDSDYDREKLQERLAKLAGGVAVIKAGAAATEVELKERKHR						

FIGURE 3 CONT'D

421	430	440	450	460	470	480	490
HUMP1	VTDALNATRAAVEGIVLGGGCALLRCIPALDSLTSPANEDQKIGIEIIKRTLKIPAMTI						
ML65K	IEDAVRNAKAAVEEGIVAGGGVTLQAAAPALDKLTLTGDEAT-GANIVKVALEAPLKQIAFN						
491	500	510	520	530	540	550	560
HUMP1	VEKIMQSSSEVGVDAMAGDFVNMVEKGIIDPTKVVRTALLDAAGVASLLTTAEVVVVTEIPKEE						
ML65K	AEKVRNLSVGHGLNAATGEYEDLLKAGVADPVKVTTRSALQNAASIAGLFTT-EAVVADKPEKTAAPASDP						
561	570						
HUMP1	MGGMGGMGGGGMF						
ML65K	TGGMGG-MD---F						

Total score = 4552, 7 breaks
 255 identities out of 540 possible matches between residues

25 random runs
 Alignment score = 47.73 SD Standard deviation = 23.86 Mean = 3413.16

FIGURE 4

	1	10	20	30	40	50	60	70
HUMP1	MLRLPTVFRQMRPVSRLAPHLTRAYAKDVKFGADARALMLQGVDDLADAVATMGPKGRTVIEQSWGS							
TB65K	M-----AKTIAYDEEARRGLERGLNALADAVKVTGLGPKGRNVVLEKKWGA							
	71	80	90	100	110	120	130	140
HUMP1	PKVTKDGVTVAKSIDLKDKYKNIGAKLVQDVANNNTNEEAGDGTATVLAARSIAKEGFEKISKGANPVEI							
TB65K	PTITNDGVSIAKEIELEDPEYKIGAEVLVKEVAKKTDVAGDGTATVLAQALRKEGLRNVAAGANPLGL							
	141	150	160	170	180	190	200	210
HUMP1	RRGVMLAVDAVIAELKKQSKPVTTPPEIAQVATISANGDKEIGNIISDAMKKVGRKGVITVKDGGKTLNDE							
TB65K	KRGIEKAVEKVTETLLKGAKEVETKEQIAATAAISA-GDQSIGDLIAEAMDKVGNVITVEESNTFGLQ							
	211	220	230	240	250	260	270	280
HUMP1	LEIEGMKFDRGYISPYFINTSKGQKCEFQDAYVLLSEKKISSIQSIVPALEIANAHRKPLVIAEDVDG							
TB65K	LELTEGMRFDKGYISGYFVTDPERQEAVLEDPYILLVSSKSVTVKDLLPLEKVIGAGKPLLIAEDVEG							
	281	290	300	310	320	330	340	350
HUMP1	EALSTLVNRLKVLQVAVKAPGFGDNRKNQKDMAIATGGAVFGEGLTNLEDVQPHDLGKVGEVIV							
TB65K	EALSTLVNKRIGTFKSVAVKAPGFGDRRKAMLQDMAITGGQVISEE-VGLTLENADLSLLGKARKVVV							
	351	360	370	380	390	400	410	420
HUMP1	TKDDAMLLKGKDKAQIEKRIQEIIEQLDVTTSEYEKEKLNERLAKLSDGVAVLKVGTSDEVNEKKDR							
TB65K	TKDETTIVEGAGDTDAIAGRVAQIRQEIENSDDYDREKLQERLAKLAGGVAVIKAGAAATEVELKERKHR							

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FIGURE 4 CONT'D

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421, 430, 440, 450, 460, 470, 480, 490,
HUMP1 VTDALNATRAAVEEGIVLGGGCALLRCIPALDSLTPANEDQKIGIEIKRTLKIPAMTIKKNAGVEGSLI
      :::::::::::::::::::::
TB65K IEDAVRNAKAAVEEGIVAGGGVTLQAAPTLDELK-LEGDEATGANIVKVALEAPLKQIAFNSGLEPGVV

491, 500, 510, 520, 530, 540, 550, 560,
HUMP1 VEKIMQSSSEVGVDAMAGDFVNMVEKGIIDPTKVVRTALLDAAGVASLLTTAEVVVTEIPKEEKDPGMGA
      :::::::::::::::::::::
TB65K AEKVRNLPAGHGLNAQTGVYEDLLAAGVADPVKVTRSAQNAAASIALGLFLTTEAVVADKPEKASVPG-

561, 570,
HUMP1 MGGMGGMGGMGMF
      ::::::
TB65K ----GGDMGGMDF

```

Total score = 4560, 5 breaks

257 identities out of 540 possible matches between residues

25 random runs

Alignment score = 49.36 SD Standard deviation = 23.23 Mean = 3413.16